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Note

Sensitive detection of unsaturated disaccharides from chondroitin sulphates by thin-layer chromatography as their dansylhydrazine derivatives

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Paper chromatography¹ has been used for the analysis of unsaturated disaccharides after the digestion of chondroitin sulphates (ChS) by chondroitinase ABC or AC. Thin-layer chromatography (TLC) on cellulose plates has also been used by many workers²⁻⁴. Recently, high-performance liquid chromatography (HPLC) has been used for the determination of unsaturated disaccharides⁵⁻¹², but TLC is still useful as a simple analytical method.

Shimada and co-workers reported the application of TLC on silica gel for the separation of hyaluronate oligosaccharides¹³ and unsaturated chondroitin sulphate disaccharides¹⁴. However, colorimetric detection on TLC plates with a carbazole reagent is not sensitive enough.

In this paper, we describe the TLC separation of small amounts of unsaturated disaccharides as their dansylhydrazine [1-naphthalenesulphonyl-5-(dimethylamino)hydrazide] derivatives, which have been used in fluorimetric precolumn derivatization in HPLC^{10,11}.

EXPERIMENTAL

Reagents and materials

The standard unsaturated disaccharides 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-D-galactose (Δ Di-0S), 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-4-O-sulpho-D-galactose (Δ Di-4S), 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-6-O-sulpho-D-galactose (Δ Di-6S), 2-acetamido-2-deoxy-3-O-(2-O-sulpho- β -D-glucopyranosyluronic acid)-D-galactose (Δ Di-UA2S), 2-acetamido-2-deoxy-3-O-(2-O-sulpho- β -D-glucopyranosyluronic acid)-4-O-sulpho-D-galactose (Δ Di-diS_B), 2-acetamido-2-deoxy-3-O-(2-O-sulpho- β -D-glucopyranosyluronic acid)-6-O-sulpho-D-galactose (Δ Di-diS_D), 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-4,6-di-O-sulpho-D-galactose (Δ Di-diS_E), 2-acetamido-2-deoxy-3-O-(2-O-sulpho- β -D-glucopyranosyluronic acid)-4,6-di-O-sulpho-D-galactose (Δ Di-triS), 2-acetamido-2-deoxy-3-O-(β -D-glucopyranosyluronic acid)-D-glucose (Δ Di-HA) and chondroitinase ABC were purchased from Seikagaku Kogyo (Tokyo, Japan) and dansylhydrazine from Fluka (Buchs, Switzerland). All other chemicals were of analytical-reagent grade.

Kieselgel 60 thin-layer plates (plastic plate, 0.2 mm thickness) were obtained from Merck (Darmstadt, F.R.G.).

A UV lamp (Model UVGL-15 Mineralight Lamp; San Gabriel CA, U.S.A.) was used to detect the spots on the developed TLC plates.

Separation of glycosaminoglycans in rabbit plasma and urine

Plasma glycosaminoglycans (GAGs) were separated according to the method of Emura and Mukuda¹⁵ with a 100- μ l sample plasma.

Urinary GAGs were separated according to Poulsen's method¹⁶.

Enzymatic digestion

Aliquots of 10 μ l each of 0.2 M Tris-HCl buffer (pH 8.0) and an aqueous solution of 0.1 U chondroitinase ABC were added to 20 μ l of plasma or a urinary GAG solution. The mixture was incubated at 37°C for 3 h and then lyophilized. The residue was dissolved in 10 μ l of water. This solution was used for dansylhydrazine derivatization.

Fluorescent derivatization of unsaturated disaccharides

Aliquots of 20 μ l of a 0.75% (w/v) solution of trichloroacetic acid in ethanol and 20 μ l of a 1.0% (w/v) solution of dansylhydrazine in ethanol were added to 10 μ l of an aqueous solution of unsaturated disaccharides produced enzymatically from plasma or urinary GAGs. The mixture was incubated at 40°C for 150 min, then cooled to room temperature. An aliquot of 5 μ l of the resulting solution was submitted to TLC.

RESULTS AND DISCUSSION

Separation of unsaturated disaccharides

The separation of dansylhydrazine derivatives of unsaturated disaccharides was examined under various conditions using commercial silica plates. It was found that Δ Di-0S, Δ Di-4S, Δ Di-6S, Δ Di-UA2S, Δ Di-diS_B, Δ Di-diS_D, Δ Di-diS_E, Δ Di-triS and Δ Di-HA could be resolved satisfactorily with *n*-propanol-isopropanol-*n*-butanol-water (30:45:5:20, v/v) containing 0.04 M sodium chloride and 0.01 M ammonia (Table I).

The detection limits of Δ Di-0S, Δ Di-4S and Δ Di-6S were 90 pmol with observation under UV light with the naked eye.

TABLE I

R_F VALUES OF DANSYLHYDRAZINE DERIVATIVES OF UNSATURATED DISACCHARIDES

<i>Unsaturated disaccharide</i>	R_F	<i>Unsaturated disaccharide</i>	R_F
Δ Di-0S	0.72	Δ Di-diS _D	0.52
Δ Di-4S	0.56	Δ Di-diS _E	0.41
Δ Di-6S	0.61	Δ Di-triS	0.30
Δ Di-UA2S	0.66	Δ Di-HA	0.75
Δ Di-diS _B	0.47		

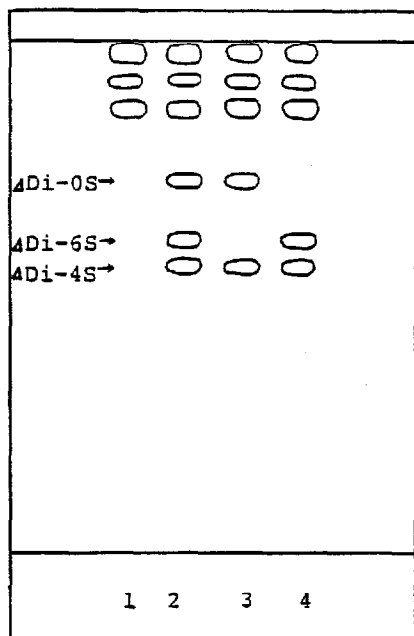


Fig. 1. Chromatogram of dansylhydrazine derivatives of unsaturated disaccharides produced from plasma and urinary chondroitin sulphates: 1 = blank; 2 = mixture of Δ Di-0S, Δ Di-4S and Δ Di-6S; 3 = plasma ChS; 4 = urinary ChS. Conditions: TLC plate, silica gel 60; developing solvent, *n*-propanol–isopropanol–*n*-butanol–water (30:45:5:20, v/v) containing 0.04 *M* sodium chloride and 0.01 *M* ammonia.

Although the TLC of Δ Di-0S, Δ Di-4S and Δ Di-6S has been studied previously^{2-4,14}, the separation of oversulphated unsaturated disaccharides such as Δ Di-diS_B, Δ Di-diS_D, Δ Di-diS_E and Δ Di-triS has not been reported. Using the method described here, all unsaturated disaccharides produced enzymatically from ChS can be separated satisfactorily with development under isocratic conditions.

Identification of unsaturated disaccharides from ChS in rabbit plasma and urine

The proposed method was applied to the identification of unsaturated disaccharides produced enzymatically from ChS in rabbit plasma and urine. Fig. 1 shows the chromatogram of dansylhydrazine derivatives of unsaturated disaccharides from rabbit plasma and urinary GAGs obtained by digestion with chondroitinase ABC.

Δ Di-0S and Δ Di-4S were detected in plasma ChS and Δ Di-4S and Δ Di-6S in urinary ChS.

In conclusion, the proposed method is useful for the identification of the unsaturated disaccharides produced from GAGs such as ChS by chondroitinases, and can be used for their structural characterization and as a screening test for ChS in biological samples because of its high sensitivity.

REFERENCES

- 1 H. Saito, T. Yamagata and S. Suzuki, *J. Biol. Chem.*, 243 (1968) 1536.
- 2 L. Wasserman, A. Ber and D. Allalouf, *J. Chromatogr.*, 136 (1977) 342.
- 3 Y. Mikuni-Takagaki and B. P. Toole, *J. Biol. Chem.*, 254 (1979) 8409.
- 4 A.-M. Säämänen and M. Tammi, *Anal. Biochem.*, 140 (1984) 354.
- 5 K. Murata and Y. Yokoyama, *Anal. Biochem.*, 146 (1985) 327.
- 6 J. Macek, J. Krajičková and M. Adam, *J. Chromatogr.*, 414 (1987) 156.
- 7 K. Murata and Y. Yokoyama, *J. Chromatogr.*, 415 (1987) 231.
- 8 T. Gherezghiher, M. C. Koss, R. E. Nordquist and C. P. Wilkinson, *J. Chromatogr.*, 413 (1987) 9.
- 9 C. Kodama, N. Ototani, M. Isemura and Z. Yosizawa, *J. Biochem.*, 96 (1984) 1283.
- 10 K. Shinomiya, I. Koshiishi, T. Imanari, M. Takeda, M. Maeda and A. Tsuji, *Chem. Pharm. Bull.*, 34 (1986) 4887.
- 11 K. Shinomiya, S. Yamanashi and T. Imanari, *Biomed. Chromatogr.*, 2 (1987) 169.
- 12 H. Toyoda, K. Shinomiya, S. Yamanashi, I. Koshiishi and T. Imanari, *Anal. Sci.*, 4 (1988) 381.
- 13 E. Shimada and G. Matsumura, *J. Biochem.*, 96 (1984) 721.
- 14 E. Shimada, N. Kudoh, M. Tomioka and G. Matsumura, *Chem. Pharm. Bull.*, 35 (1987) 1503.
- 15 Y. Emura and T. Mukuda, *Seikagaku*, 45 (1973) 30.
- 16 J. H. Poulsen, *Scand. J. Clin. Lab. Invest.*, 41 (1981) 367.